

MICROBIOLOGY CLINICAL MENTORSHIP



VM 21300

CRITERIA HANDBOOK AND LOGBOOK

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NOTE THE FOLLOWING DUE DATES FOR THE TASKS ABOVE:

Fall or Spring semester 5:00p.m. Thursday of week 8 – Tasks 1-8

Summer session 5:00p.m. Thursday of week 5 – Tasks 1-8

Incomplete grades will not be assigned for mentorships at the end of the semester.

Grade penalties will be assessed for tasks submitted after the due date.

Resubmission due dates will be set by the instructor as required.

STUDENT INFORMATION

GOALS OF VM 21300

MICROBIOLOGY CLINICAL MENTORSHIP

Working with a small animal veterinary care facility, the student will practice several tasks under the supervision of a clinical mentor (veterinarian or Credentialed Veterinary Technician).

In order to achieve the goals for this Clinical Mentorship, the tasks must be performed to the level of competency as outlined by the *Criteria* for each task.

The student is responsible for providing documentation for each task as defined by the *Materials Submitted for Evaluation and Verification* section on each task.

In addition to the documentation, the Clinical Mentorship site supervisor will verify that the student performed the task under their supervision.

Final approval of successful performance and completion of the Clinical Mentorship will be made by the Purdue University instructor in charge of the Clinical Mentorship. This approval will be based upon the documentation provided by the student.

The Purdue University instructor in charge has the option to require additional documentation if, in their judgment, the student has not performed and/or documented the task to the level set by the Criteria.

Documentation of completed tasks is essential to validate the educational process and insure that the performance of graduates of the Veterinary Technology Distance Learning Program meets the standards of quality required by the Purdue University College of Veterinary Medicine faculty and the American Veterinary Medical Association accrediting bodies.

CONTACT PERSON

Any questions regarding the Clinical Mentorship process should be directed to:

Pam Phegley, BS, RVT
Purdue University
Veterinary Technology Program
625 Harrison Street, Lynn Hall G171
West Lafayette IN 47907
(765) 496-6809
phegleyp@purdue.edu

PRE-REQUISITES FOR VM 21300 MICROBIOLOGY CLINICAL MENTORSHIP

Contracts and Agreements

Because of legal, liability and AVMA accreditation issues, the following documents must be completed *prior to beginning* the Clinical Mentorship

1. Facility Requirement Agreement
2. Clinical Mentorship Agreement
3. Supervisor Agreement
4. Health Risk and Insurance Acknowledgement
5. Professional Liability Insurance Coverage
6. Agreement and Release of Liability
7. Technical Standards Acknowledgement
8. Code of Conduct

These forms are available on the VTDL website for downloading, printout, and completion.

If more than one Clinical Mentorship course is taken, a separate Facility Requirement Agreement, Clinical Mentorship Agreement, and Supervisor Agreement must be completed for each course.

More than one Mentorship Supervisor may sign the mentorship logbook. Each must be either a DVM or a credentialed technician, and must complete a separate Supervisor Agreement.

Failure to complete and return the listed documents and the payment for Student Professional Liability Insurance Coverage will prevent the student from enrolling in the Clinical Mentorship.

Insurance

Two types of insurance are recommended or required for the student working in a Clinical Mentorship.

Health Insurance is highly recommended to cover the medical expenses should the student become injured while on the job. It is the student's responsibility to procure such insurance.

Liability Insurance is required to protect the student in the event of a suit filed against the student for acts he/she performed while in the Clinical Mentorship.

Each VTDL student is required to purchase, for a nominal fee, Professional Liability Insurance through Purdue University. This is done by completing the Professional Liability Insurance Coverage form and sending a check for the fee. This check must be separate from payment of course fees. The fee covers from the time of initiation of coverage until the subsequent July 31st.

Students will not be enrolled in Clinical Mentorships until the Professional Liability Insurance is paid, and the student is covered by the policy.

SELECTING THE CLINICAL MENTORSHIP SITE – FACILITY REQUIREMENTS

You must visit the Clinical Mentorship Site and determine if the following supplies and equipment are readily available to you for use during your Clinical Mentorship. You must complete and have the facility veterinarian sign the Facility Requirement Agreement.

The veterinary care facility must be equipped with the following equipment:

- Forceps or hemostats
- Ruler with mm markings
- Flame source (cigarette lighter, wooden matches, Bunsen burner)
- Microscope with 10X, 40X, 100X
- Incubator

The veterinary care facility must be equipped with the following items:

- Cotton balls
- 70% isopropyl alcohol
- Scalpel blades (#10 or 15)
- Sterile cotton-tipped applicators
- Sterile saline
- Microscope slides
- Immersion oil
- 3% hydrogen peroxide
- Wax pencil or crayon

The student will be provided with a kit which will include:

- Disposable inoculating loops
- Disposable inoculating needles
- Disposable pipettes
- Gram stain kit
- Blood agar plates
- Nutrient broth tubes
- Mueller-Hinton agar plates
- Dermatophyte test medium
- Oxidase reagent
- 0.5 McFarland nephelometer
- Antibiotic discs

NOTE: The student will be informed before the kit is sent, and materials must be used within one week in order to obtain accurate results. It is imperative that the student follow all steps closely for these tasks, as additional materials will not be sent. If there are questions please contact the instructor, Josh Clark, at clark19@purdue.edu BEFORE performing the task.

SELECTION OF THE CLINICAL MENTORSHIP SUPERVISOR

The Clinical Mentorship Supervisor is the person who will sign your Logbook and verify performance of tasks at the Clinical Mentorship site. This person must be a credentialed veterinary technician (have graduated from an AVMA accredited program or met State requirements for credentialing as a veterinary technician) or a licensed veterinarian.

An individual who claims to be a “veterinary technician” but has not met the criteria for credentialing above is not eligible to be mentorship supervisor.

The individual is not considered to be an employee of Purdue University when acting as your Clinical Mentorship supervisor.

Each Clinical Mentorship Supervisor must complete a *Supervisor Agreement*. You must return this agreement with the other agreements prior to beginning your Clinical Mentorship. Multiple supervisors may be used for documentation of mentorship tasks. Each supervisor must complete a separate agreement.

Should your Clinical Mentorship Supervisor change during the course of the Clinical Mentorship, you will need to have your new supervisor complete a *Clinical Mentorship Supervisor Agreement* and return it to the Purdue VTDL office. These forms are available on the VTDL website for downloading and printing.

CRITERIA HANDBOOK AND LOGBOOK

This Criteria Handbook and Logbook contains the list of tasks that must be successfully completed in order to receive credit for this Clinical Mentorship. You are expected to have learned the basics of how, why, and when each procedure is to be done from the courses listed as pre-requisites for this Clinical Mentorship. This booklet contains the directions and forms that must be followed and completed in order to meet the standards set for successful completion of this Clinical Mentorship.

Please read each component of each task carefully before doing the task to minimize the number of times you have to repeat the task. The components of each task are summarized:

Goal – Describes the ultimate outcome of the task you will perform.

Description – Lists the physical acts that you will perform, and under what conditions these acts will be completed.

Criteria – Lists specific, observable, objective behaviors that you must demonstrate for each task. Your ability to demonstrate each of these behaviors will be required in order to be considered as having successfully completed each task.

Number of Times Task Needs to be Successfully Performed – States the required number of times to repeat the tasks. The patient's name and the date each repetition of the task was performed must be recorded on the Task Verification Form.

EACH REQUIRED REPETITION OF THE TASK MUST BE PERFORMED ON A DIFFERENT ANIMAL. You cannot use the same animal to do all of the repetitions of a task. However, you can use the same animal to perform different tasks. In other words, you can't do three ear cleanings on the same animal, however, you can do an ear cleaning, an anal sac expression, and a venipuncture on the same animal.

Materials Submitted for Evaluation and Verification – These specific materials, which usually include video or other materials, must be submitted to demonstrate that you actually performed the task as stated. Each evaluation states specifically what must be shown in the submitted materials.

The Purdue University course instructor for this Clinical Mentorship has the option to request further documentation if the submitted materials do not clearly illustrate the required tasks.

It is recommended that the video materials document all angles of the procedure. The purpose of the video and other material is to provide "concrete evidence" that you were able to perform the task to the standard required.

If you do not own a video camera, one may be borrowed or rented. Pre-planning the video procedures will help reduce the need to redo the video documentation. Explain what you are doing as you perform the video documentation, as narration will help the evaluator follow your thought process and clarify what is seen on the video. Voiceovers may be done to clearly explain what is being performed. At the beginning of each task, clearly announce what task you are doing, or insert a written title in the video.

Videotapes, photographs, radiographs, slides, written projects, the Criteria Handbook and Logbook and any other required documentation will not be returned. These items will be kept at Purdue as documentation of the student's performance for accreditation purposes.

This validation is essential to help the Purdue VTDL meet AVMA accreditation criteria. Therefore, it is essential that you follow the evaluation and validation requirements.

Task Verification Forms – Each task has a form that must be completed and signed by the Clinical Mentorship Supervisor.

Supplementary Materials – Logs, written materials, photographs, or other forms/documentation may be required for specific tasks. Be sure to read the Materials to be Submitted for Evaluation section very carefully and return all documented evidence as prescribed.

COMPLETION OF THE CLINICAL MENTORSHIP

Mentorship logbooks include due dates for sections of courses. Each section must arrive at Purdue by the deadline (not a postmark date).

Paperwork may be

- FAXed to 765-496-2873
- e-mailed to phegleyp@purdue.edu
- sent by regular mail to 625 Harrison Street, Lynn Hall G171, West Lafayette, IN 47907

Videos may be submitted

- in the Media Gallery of Blackboard. If submitted on Blackboard, send an e-mail to phegleyp@purdue.edu notifying of the submission. ***This is the preferred method of online submission***, since it does not limit how much you put on, is no cost to you, and automatically archives here. You must assign the videos to the correct course in order for the instructor to view them.
- by an online source such as Dropbox. If a password is required to open videos submitted with an online service, email the password to phegleyp@purdue.edu. These methods may not be acceptable if they cannot be archived.
- by sending on a disc or flash drive by regular mail to 625 Harrison Street, Lynn Hall G171, West Lafayette, IN 47907

Late submissions will incur a grade penalty. Incomplete grades will no longer be assigned for mentorships at the end of each semester.

Feedback will be emailed until all tasks are completed successfully. A hard copy will be sent when the course is complete and a grade is assigned. As necessary, instructors may require resubmission of some tasks. When feedback is sent, due dates for resubmissions will be given. *It is crucial that students with pending feedback check their Purdue emails frequently so this information is received in a timely manner.*

Final approval of successful performance and completion of the Clinical Mentorship will be made by the Purdue University instructor in charge of the Clinical Mentorship based upon the documentation provided by the student.

Upon successful completion of all tasks in the clinical mentorship course, a grade will be assigned by the course instructor based upon the documented performance of the tasks.

CLINICAL MENTORSHIP TASKS

INTRODUCTION TO ESSENTIAL TASKS AND CRITERIA

Before starting each task:

1. Read the Goal, Description, Criteria, and Materials to be Submitted for Evaluation and Verification. Understand what is expected of you for each task.
2. Make sure you have whatever equipment and supplies you need to document the task. Pay particular attention to the details of what needs to be documented and submitted.
3. Make sure you obtain appropriate permissions where necessary. Please inform the facility's owner/manager of your activities. A good relationship with the veterinarian in charge is key to having a positive Clinical Mentorship experience.

After performing each task:

4. Label all items submitted so that the materials you submit for evaluation and validation at Purdue are identified as your submission.
5. Label all videos posted to Blackboard with the name of the task performed.
6. Submit materials to Purdue by the deadlines listed in the logbooks.

CLINICAL MENTORSHIP PROJECTS

INTRODUCTION TO SPECIAL PROJECTS

Certain mentorships will have required projects to complete in addition to the required tasks. These are things that are better assessed in the form of a project. Projects should be typed, and checked for correct grammar and spelling.

Before starting each project

1. Read through the project in its entirety. This will give you a description of the project and what is needed to complete it successfully.
2. Determine what materials, if any, need to be submitted for completion of the project.
3. Most projects will come with a list of questions that need to be answered. The responses should be placed inside the notebook for submission with other materials.
4. If videotaping is required for a project, it should be noted on the videotape verbally that this is for the project and not another required task. Some projects may require a verbal narration of a student doing something. Each individual project will define if that is a necessary requirement for that project.

Note: Videotaping and photographs are not for the purpose of verifying if the practice is within OSHA compliance or other government regulations. These projects are for the student's education. It may be determined by the student that the practice is not within the current recommendations. The purpose of these projects is to make the student aware of these issues, and how to recognize the issues and develop suggestions for improvement. There will be certain mentorships where OSHA recommendations, in regards to equipment and policies, will be facility requirements for the mentorship.

****Important Information about Performing Microbiology Tasks****

Before your supplies arrive, we highly recommend that you go into the VM 22200 course in Blackboard to review the instructional videos. You should also review the task criteria in the VM 22200 logbook. If you have any questions about successfully completing the criteria for each of the microbiology tasks, contact the instructor for clarification. You will need to allow seven consecutive days for performance of the microbiology tasks. Following is an outline of what should be done each day:

Day 1

- Collect samples and inoculate dermatophyte culture media (Derm-Duet)
- Swab several things (or animals) in the clinic and streak several blood agar plates to obtain isolated colonies
- Collect a urine sample and inoculate a tube of nutrient broth

Day 2

- Select isolated colonies from the blood agar plates that you inoculated on day one and streak several new blood agar plates to obtain isolated colonies in a pure culture (i.e., only one type of bacteria growing on the plate)
- Check the nutrient broth you inoculated with urine for any signs of growth
- Check dermatophyte culture for growth and color change

Day 3

- Utilize well-isolated colonies from the blood agar plates that you inoculated on day two to create your gram stain slides
- Utilize well-isolated colonies from the blood agar plates that you inoculated on day two to inoculate your Mueller-Hinton plates, and place antimicrobial discs on the plates
- Utilize well-isolated colonies from the blood agar plates that you inoculated on day two to perform your oxidase and catalase tests
- Check the nutrient broth you inoculated with urine for any signs of growth
- Check dermatophyte culture for growth and color change

Day 4

- Interpret and record the results of your Kirby Bauer test
- Check dermatophyte culture for growth and color change

Days 5-6

- Check dermatophyte culture for growth and color change

Day 7

- Show your dermatophyte culture on your video and verbally describe the appearance of any growth as well as any changes to the color of the agar

1. COLLECT A SAMPLE FOR DERMATOPHYTE CULTURE AND INOCULATE DERMATOPHYTE CULTURE MEDIA

Goal: To collect an adequate sample for Dermatophyte culture and use that sample to inoculate Dermatophyte Culture Media in a manner that will yield accurate results.

Description: The student will identify a possible Dermatophyte lesion and scrape/pluck an appropriate sample from the edge of the lesion. The student will use the sample to inoculate the Dermatophyte Culture Media.

Criteria: The student identified a suspected Dermatophyte lesion

The student washed and dried the affected area with soap and water

The student obtained a small scraping of superficial debris and hair from the margin of the lesion using a sterile scalpel blade and forceps, or plucked a sample of hair from the margin of the lesion using hemostats

The student inoculated the culture media by placing the sample slightly below the surface of the media

The student left the lid to the tube or plate slightly open

The student allowed the culture to incubate at room temperature

The student showed the plate after at least seven days' incubation and verbally stated whether the result was positive or negative at that time (understanding that the plate should be checked daily for up to one month)

Number of Times Task Needs to be Successfully Performed: 2

Materials Submitted for Evaluation and Verification:

1. Task verification form for Collect a Sample for Dermatophyte Culture and Inoculate Dermatophyte Culture Media task, signed by the Clinical mentorship supervisor.
2. A video that clearly shows the student collecting a sample for Dermatophyte culture and inoculating Dermatophyte culture media, AND showing growth/results as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Patient Name: _____ **Date:** _____

Patient Name: _____ **Date:** _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

2. COLLECT SWAB FOR CULTURE & STREAK AN AGAR PLATE TO OBTAIN ISOLATED COLONIES

Goal: To collect an adequate sample for culture and use that sample to inoculate an agar plate in a manner that will yield discrete isolated colonies of growth.

Description: The student will collect a sample with a sterile swab from an area in the hospital or from a patient that would likely yield bacterial growth. The student will use the sample to inoculate a blood agar plate.

Criteria: The student lightly moistened a sterile swab with sterile saline

The student used the sterile swab to collect a sample from an area of the hospital (e.g. dirty sink, floor, drain, etc.) or from a patient (nose, mouth, ear, wound) and used that swab to apply the primary streak on a blood agar plate

The student rotated the plate and used an inoculating loop to streak the next quadrant of the blood agar plate, being careful to overlap the primary streak only once or twice

The student rotated the plate and used an inoculating loop to streak the third quadrant of the blood agar, being careful to overlap the second streak only once or twice

The student rotated the plate again and used an inoculating loop to streak the fourth and final quadrant of the blood agar, being careful to overlap the third streak only once or twice

The student inverted the plate and placed it in an incubator for 18-24 hours

The student checked the plate after the incubation period for isolated colonies of growth, showing the plate and verbally stating observations (e.g., colony characteristics)

Number of Times Task Needs to be Successfully Performed: 2

Materials Submitted for Evaluation and Verification:

1. Task Verification form for Collect Swab for Culture & Streak an Agar Plate to Obtain Isolated Colonies task, signed by the Clinical Mentorship supervisor.
2. A video that clearly shows the student collecting a sample swab and streaking a blood agar plate using the streak isolation technique, AND showing growth as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Patient Name: _____ **Date:** _____

Patient Name: _____ **Date:** _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

3. STREAK AGAR PLATE TO OBTAIN A PURE CULTURE

Goal: To collect a sample from an isolated colony and streak an agar plate to obtain a pure culture with isolated colonies.

Description: The student will select an isolated colony of bacteria with an inoculating loop and streak it onto a blood agar plate using the streak isolation technique to obtain a pure culture with isolated colonies.

Criteria: The student used an inoculating loop to collect a sample of bacteria from an isolated colony from a blood agar plate inoculated for the previous task, and used that sample to apply the primary streak on a blood agar plate.

The student rotated the plate and used an inoculating loop to streak the second quadrant of the blood agar, being careful to overlap the primary streak only once or twice

The student rotated the plate and used an inoculating loop to streak the third quadrant of the blood agar, being careful to overlap the second streak only once or twice

The student rotated the plate and used an inoculating loop to streak the fourth and final quadrant of the blood agar, being careful to overlap the third streak only once or twice

The student inverted the agar plate and placed it in an incubator for 18-24 hours

The student checked the plate after the incubation period for isolated colonies of growth, showing the plate and verbally stating observations

Number of Times Task Needs to be Successfully Performed: 10

Materials Submitted for Evaluation and Verification:

1. Task Verification form for Streak Agar Plate to Obtain a Pure Culture task, signed by the Clinical Mentorship supervisor.
2. A video that clearly shows the student collecting a sample from an isolated colony with an inoculating loop and streaking the plate using the proper technique, AND showing growth as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Date: _____ Date: _____ Date: _____ Date: _____ Date: _____

Date: _____ Date: _____ Date: _____ Date: _____ Date: _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

4. COLLECT A URINE SAMPLE FOR CULTURE & INOCULATE NUTRIENT BROTH

Goal: To collect an appropriate sample of urine and use the sample to inoculate nutrient broth.

Description: The student will select an appropriate collection device and collect at least 1ml of urine by cystocentesis, catheterization or free catch. Note: Cystocentesis is generally the only acceptable method of collection of urine for culture; however for this exercise the student may collect the urine sample by other, less invasive methods. The sample will then be used to inoculate nutrient broth. If the urine is collected by free catch, the collection container must be sterile.

Criteria: The student prepared the collection site as needed, depending on the collection method, and collected the urine sample into an appropriate container (syringe and needle, or sterile container).

The student removed the cap of the nutrient broth tube with the last two fingers

The student dipped the sterile inoculating loop into the urine sample

The student inserted the loop into the sterile nutrient broth and inoculated it by gently moving the loop back and forth

The student removed the inoculating loop from the tube and replaced the cap on the tube

The student placed the nutrient tube into the incubator for 24 hours and then checked it for growth, showing the tube and verbally stating observations

The student placed the nutrient tube back in the incubator for another 24 hours and then checked it for growth, showing the tube and verbally stating observations

Number of Times Task Needs to be Successfully Performed: 2

Materials Submitted for Evaluation and Verification:

1. Task Verification form for Collect a Urine Sample for Culture & Inoculate an Nutrient Broth task, signed by the Clinical Mentorship supervisor.
2. A video that clearly shows the student collecting a urine sample for culture and nutrient broth in a tube, AND showing growth at 24 and 48 hours, as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Patient Name: _____ **Date:** _____

Patient Name: _____ **Date:** _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

5. PERFORM KIRBY-BAUER DISC SENSITIVITY TEST

- Goal:** To successfully perform a Kirby-Bauer Disc Sensitivity Test such that one can measure zones and determine the bacterial sample's resistance or susceptibility to certain antibiotics.
- Description:** The student will perform a Kirby-Bauer Disc Sensitivity Test on an organism obtained from an isolated colony in such a way that the bacterial sample can be determined as being susceptible, resistant, or intermediate to certain antibiotics.
- Criteria:**
- The student selected 4-5 morphologically identical colonies from a blood agar or MacConkey plate and aseptically transferred the selected colonies to a tube of sterile saline with an inoculating loop
 - The student mixed the contents of the tube by swirling (not inverting) the tube so that the organisms were uniformly suspended in the saline
 - The student compared their tube with 0.5 McFarland nephelometer to ensure that the turbidity in the tube was the same
 - The student adjusted the turbidity in their tube as needed, by dilution or adding of colonies, to attain a suspension of bacteria with a turbidity equal to the 0.5 McFarland nephelometer
 - The student used aseptic technique to dip a sterile cotton swab into the saline suspension of bacteria
 - The student rotated the swab against the inner wall of the tube to express excess fluid
 - The student swabbed the **entire surface** of a Mueller-Hinton agar plate with the swab
 - The student rotated the plate 60° and reswabbed the **entire surface**
 - The student again rotated the plate 60° and reswabbed the **entire surface**, then ran the swab around the outside edge of the agar
 - The student applied the antibiotic discs with flamed forceps that were allowed to cool between uses. The student gently pressed each disc on the agar surface to ensure complete contact, and made sure the discs were at least 24 mm apart from center to center. The student also ensured that the discs did not move after contact with the agar surface was made
 - The student allowed the plate to sit for 1-2 minutes, then inverted the plate and placed it into an incubator
 - The student removed the plate from the incubator after 16-18 hours and showed the plate, verbally stating observations
 - The student measured the cleared zone **diameters** in mm, through the bottom of the plate
 - The student recorded the measurements and stated them verbally
 - The student consulted the lab table located in this logbook to determine the susceptibility character of the organism, and verbally stated the results

Number of Times Task Needs to be Successfully Performed:
Materials Submitted for Evaluation and Verification:

2

1. Task Verification form for Perform a Kirby-Bauer Disc Sensitivity Test task, signed by the Clinical Mentorship supervisor.
2. One video that clearly shows the student performing Kirby-Bauer Disc Sensitivity Test, AND showing growth, measurement and results as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Date: _____

Date: _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

6. PREPARE A GRAM-STAINED SLIDE

Goal: To prepare a Gram-stained slide such that one is able to look at the slide under the 100X oil immersion lens of a microscope and identify morphology and Gram reaction of the bacteria on the slide.

Description: The student will prepare a Gram-stained slide with an organism obtained from an isolated colony in such a way that the morphology and Gram reaction of the organism on the slide may be identified.

Criteria: The student selected a bacterial sample by touching a sterile wire or loop to one colony on an agar plate (did not scoop an entire colony off the agar plate)

The student mixed the sample on a microscope slide with a drop of water or saline

The student circled the sample droplet on the slide with a wax pencil to help identify the area after staining

The student allowed the slide to air dry

The student heat fixed the slide by passing it through a flame 2-3 times, specimen side up

The student held the slide over the sink or placed it on a rack over the sink, flooded the smear with crystal violet, and let stand for one minute

The student rinsed the smear briefly with water

The student held the slide over the sink and flooded the smear with Gram's iodine solution and let stand for one minute

The student rinsed the smear briefly with water

The student washed the smear with decolorizer until no more purple color washed off (5-10 seconds)

The student rinsed the smear briefly with water

The student held the slide over the sink and flooded the smear with safranin and let stand for one minute

The student rinsed the smear briefly with water

The student allowed the slide to air dry or gently blotted it dry between paper towels

The student mounted the slide on the microscope and focused on the smear beginning with the low power lens and working up to the oil immersion lens, and verbally stated Gram Reaction and morphology

Number of Times Task Needs to be Successfully Performed: 5

Materials Submitted for Evaluation and Verification:

1. Task Verification form for Prepare a Gram Stained Slide task, signed by the Clinical Mentorship supervisor.

2. A video that clearly shows the student preparing and evaluating a Gram stain as defined in the above criteria for this task.
3. **One microscope slide** prepared by the student using the technique described for the Prepare a Gram-Stained Slide task. The slide should be clearly labeled and identified as Gram-positive or Gram-negative as well as stating morphology of the sample (rods, cocci, etc.)

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Date:_____ Date:_____ Date:_____ Date:_____ Date:_____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

7. PERFORM CATALASE TEST

- Goal:** To successfully perform a catalase test such that one is able to look at the slide and determine whether a bacteria is catalase positive or negative.
- Description:** The student will perform a catalase test on an organism obtained from an isolated colony in such a way that the sample may be determined to be catalase positive or negative.
- Criteria:** The student selected a bacterial sample by touching a sterile loop or wire to the center of one colony on an agar plate. If a blood agar plate was used, the student avoided contacting the agar with the loop or wire
- The student applied a drop of 3% hydrogen peroxide onto a microscope slide
- The student smeared the sample in the drop of hydrogen peroxide
- The student immediately observed the slide for bubbling. If using a loop, the student observed the loop for bubbling as well
- The student showed the slide and verbally identified whether the sample was catalase positive or negative

Number of Times Task Needs to be Successfully Performed: 2

Materials Submitted for Evaluation and Verification:

1. Task Verification Form for Perform Catalase Test skill, signed by the Clinical Mentorship supervisor.
2. A video that clearly shows the student performing a catalase test as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Date: _____

Date: _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____

8. PERFORM OXIDASE TEST

Goal: To successfully perform an oxidase test such that one is able to look at the swab and determine whether a bacteria is oxidase positive or negative.

Description: The student will perform an oxidase test on an organism obtained from an isolated colony in such a way that the sample may be determined to oxidase positive or negative.

Criteria: The student placed a drop of oxidase reagent onto a sterile swab

The student touched the moistened swab to an isolated colony on an agar plate

The student waited 30-60 seconds for a color change on the swab

The student showed the swab and verbally identified whether the sample was oxidase positive or negative

Number of Times Task Needs to be Successfully Performed: 2

Materials Submitted for Evaluation and Verification:

1. Task Verification Form for Perform Oxidase Test skill, signed by the Clinical Mentorship supervisor.
2. One video that clearly shows the student performing an oxidase test as defined in the above criteria for this task.

Student Name: _____

Supervisor Name: _____ RVT, CVT, LVT
DVM, VMD

Date: _____

Date: _____

I verify that the student performed these tasks under my supervision.

Signature of Clinical Mentorship Supervisor: _____